

# NATURAL COAGULANTS: AN EASY WAY TO REMOVE HEAVY METALS FROM TANNERY EFFLUENTS

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## ABSTRACT

The seeds of *Strychnos potatorum* and *Moringa oleifera* are a natural coagulant which shows promising biofloculant activity. These seeds are used to purify the drinking water in rural South India. In this research, an attempt has been made to analyze the biosorption of heavy metals by *Strychnos potatorum* and *Moringa oleifera* seeds from the tannery effluent. Heavy metals were estimated by atomic absorption spectrophotometer. The toxicity level of the treated effluent was analysed by green gram germination bioassay. Shoot length and root length of the germinated seeds were measured and seedling vigour index were calculated. The heavy metal removal efficiency of the seeds of *Moringa* and *Strychnos* was higher there by making both the seeds a promising candidate for industrial effluent treatment. These agro based compound could replace the chemical coagulant and is an environmentally friendly approach to effluent treatment.

## INTRODUCTION

The use of natural coagulants in the water treatment was focused as an alternative to the expensive chemical coagulants. Chemical coagulants such as alum, ferric chloride, calcium carbonate, polyaluminium chloride and polyethylene amine were used as common inorganic coagulants in the water treatment units. These methods include electrochemical precipitation, ultrafiltration, ion exchange and reverse osmosis. A major drawback with precipitation is sludge production (Shetie Gatew et al., 2013). The residual of alum and polyaluminium chloride may produce the Alzheimer's disease and some of the synthetic organic polymers cause the neurotoxic and carcinogenic effect (Ndabigengesere 1995). Therefore, an attempt was made to use agro based coagulants as an alternate for chemical flocculants. The agro products would be more favorable of their low cost, abundance, environment friendly and does not leave toxic residues. Moreover, cheap and illiterate labor could be employed while treating the toxic tannery effluent in large scale.

The use of natural material of plant origin to clarify turbid surface water is not a new idea. The use of

herbal materials to reduce turbidity or muddiness in the water and to remove the harmful biological material that can lead to illness is an age-old concept (Joshua and Vasu., 2013).

Plants such as *Moringa oleifera*, *Strychnos potatorum*, *Jatropha curcas*, *Hibiscus sabdariffa*, *Clidemia aungustifolia*, *Cactus opuntia*, *Phaseolus vulgaris*, *Prosopis juliflora*, *Ipomea dasysperma* and Gum Arabic, *Guar gum* as coagulants have been used for water purification. Among these *S. potatorum* (Loganiaceae) and *M. oleifera* (Moringaceae) have shown as promising coagulant in the clarification of turbid water (Tripathi et al., 1976; Sutherland et al., 1990; Folkard and Sutherland., 2002). By reviewing the literature, it was observed that *S. potatorum* and *M. oleifera* seeds have natural coagulation property as well as used as an adsorbent to remove the heavy metals from aqueous solution (Jayaram et al., 2009; Joseph et al., 2015) reported that *M. oleifera* seeds and leaves as adsorbents shows the high extraction efficiency of the metals like Na, Fe, and Zn. The seeds of *S. potatorum* contained galactan and galactomannan which reduced the Cr (VI) from aqueous solution (Muthuraman and Sasikala, 2015). The surface

modified *S. potatorum* seeds show higher adsorption capacity towards the removal of cadmium ions from aqueous solution (Senthilkumar et al., 2012). Hong et al., (2017) reported that a seed of *M. oleifera* has proved to be very effective in the removal of heavy metal like Cu, Cd, Zn, Ni, Mn and Fe in domestic wastewater.

Biosorption is a technically efficient, commercially viable and eco- friendly technology and it can be defined as the ability of biological materials to accumulate heavy metals from wastewater through metabolically mediated/physico-chemical pathways of uptake (Fourest and Roux., 1992). Agro waste contains large quantity of polysaccharides and proteins with many functional groups such as carboxyl, hydroxyl, sulphate, phosphate and amino groups, which can bind metal ions. The performance of a biosorbent depends on its surface characteristics, which can be changed by modification process (Abdulsalam et al., 2014). The seeds of *S. potatorum* extracts are anionic polyelectrolytes which destabilize the particles present in the water by interparticle bridging, whereas the *M. oleifera* seeds contain active coagulant protein called a dimeric protein that have been used for various water treatment. The efficacy of these seeds was not examined for the biosorption of metals from tanning effluent were not studied elaborately. So far, the natural coagulant was used in the treatment of dye effluents.

The usage of biomass for the removal of heavy metals from the tannery effluent has not been given adequate attention. The aim of the present study is to examine the biosorption efficiency of *S. potatorum* and *M. oleifera* seeds by treating with tannery effluent and to analyze the uptake of heavy metals by them.

## MATERIALS AND METHODS

- Collection of tannery effluent at Leather Tanning Industry, Vaniyambadi, Tamil Nadu, India.

- Collection of seeds at local Siddha Medical Shop, Ambattur, Chennai, Tamil Nadu, India.

### Preparation of samples

The seeds were ground and sieved using 0.05 mm sieve and used as an adsorbent for the removal of heavy metals from the tannery effluent.

### Physico-chemical analysis of tannery effluent:

Physicochemical parameters of raw tannery effluent such as pH, Total Dissolved Solids (TDS), Total Solids(TS), Total Suspended Solids (TSS), Total Hardness as CaCO<sub>3</sub> equ, Cl<sup>-</sup>, SO<sub>4</sub><sup>2-</sup>, Cr<sup>3+</sup>, Mg, Fe, P, COD, BOD were estimated using standard methods.

### Treatment of raw effluent using *S. potatorum* and *M. oleifera* seeds powder:

The Jar test experiment for treating effluent using natural coagulants was done. It was carried out by adding 10 gm/L of the seed powder to 1 L of tannery effluent separately and water as control. This study consists of occasional mixing, slow mixing and sedimentation. The set up was maintained for 7d at room temperature. The suspensions were allowed to settle down and separated by centrifugation (5000 × g). The supernatant and pellets were collected separately for analysis for heavy metal by Atomic Absorption Spectrophotometer (AAS). The weight of the pellet collected was determined and tabulated (Fig. 1).

**Heavy metal analysis:** The concentration of heavy metals in the tannery effluent before and after treatment and also the seed powder was analyzed using AAS. All the samples were acid digested for AAS analysis. The Atomic absorption spectrophotometer was heated with cathode lamp, the air acetylene flame was ignited and the instrument was calibrated with different working standards. The metals namely Cr, Cd, Zn, Fe, and Mg were detected in AAS (Elico instrument model SL 243 double beam) using respective standards. The biosorption percentage was calculated.

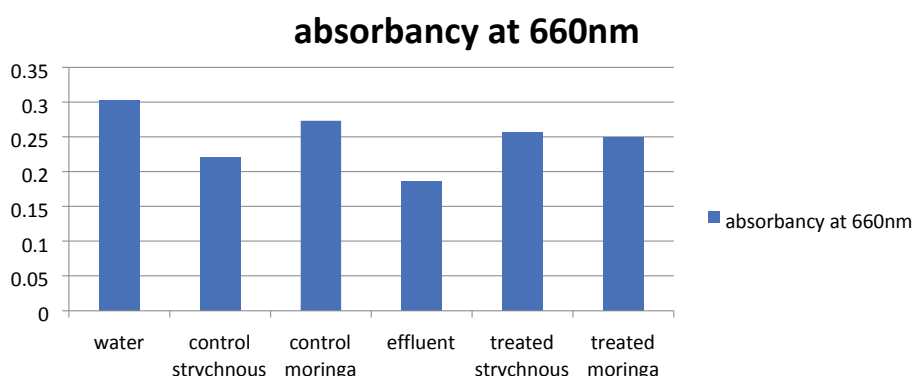


Fig 1. UV- Visible spectrophotometer analysis before and after treatment.

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The percentage of removal and the metal ion sorbent amount onto the biosorbent was calculated as (Reddy et al., 2012):

$$\text{Biosorption potential} = (C_o - C_e) / C_e \times 100$$

Where  $C_o$  and  $C_e$  are concentration of the metal ion in the solution initially and after biosorption.

**Bioassay:** The germination of Vigna radiate seeds in treated tannery effluent was observed for 3 days and root length and shoot length were measured. Germination percentage and seedling vigour index was calculated as:

$$\text{Germination\%} = (n/N) \times 100$$

$N$  = No. of seeds introduced

$n$  = No. of seeds germinated

$$\text{Seedling Vigour Index} = (\text{Shoot length} + \text{Root length}) \times \text{Germination\%}$$

## RESULTS AND DISCUSSION

### Physico - chemical analysis of tannery effluent

The physico-chemical parameters of tannery effluent such as pH (8.14), Total Dissolved Solids (10989 ppm), Total Suspended Solids (29 ppm), Cl<sup>-</sup> (5680 ppm), Cr (8 ppm), Mg (349 ppm), COD (8569 ppm), BOD (3147 ppm) are presented in Table 1.

### Effect of seed powder in the treatment of tannery effluent

Tannery effluent was treated with seed powder of *S. potatorum* and *M. oleifera* separately for 7 days. The treated effluent was separated by centrifugation at 2268

5000 × g. The pellets of treated seed powder and the treated effluent were estimated by AAS. The heavy metals namely Cr, Cd, Zn, Fe, and Mg were reported. From the Tables 2 and 3, it was found that the removal of heavy metal from the effluent was more by *S. potatorum* when compared with *M. oleifera*. *S. potatorum* seeds shows high biosorption potential of heavy metals in the order Cr > Cd > Mg > Fe > Zn and *M. oleifera* seeds in the order Cr > Zn > Fe > Mg > Cd. Biosorption of seed powder of both plants was represented in Table 4. Both the seeds biosorb Cr the most, whereas Zn was biosorbed least by *S. potatorum* while *M. oleifera* biosorbed the entire Zn ions from the effluent.

| S. No. | Parameters                                 | Experimental value |
|--------|--------------------------------------------|--------------------|
| 1      | pH                                         | 8.14               |
| 2      | Total Dissolved Solids                     | 10989 ppm          |
| 3      | Total Solids                               | 11088 ppm          |
| 4      | Total Suspended Solids                     | 29 ppm             |
| 5      | Chloride as Cl <sup>-</sup>                | 5680 ppm           |
| 6      | Sulphate as SO <sub>4</sub> <sup>2-</sup>  | 3148 ppm           |
| 7      | Chromium as Cr <sup>3+</sup>               | 8 ppm              |
| 8      | Total Hardness as CaCO <sub>3</sub> equ.   | 2146 ppm           |
| 9      | Calcium as Ca                              | 324 ppm            |
| 10     | Magnesium as Mg                            | 349 ppm            |
| 11     | Total Alkalinity as CaCO <sub>3</sub> equ. | 518 ppm            |
| 12     | Iron Content                               | 0.05 ppm           |
| 13     | Phosphate                                  | Traces             |
| 14     | Chemical Oxygen Demand                     | 8569 ppm           |
| 15     | Biological Oxygen Demand                   | 3147 ppm           |

**Table 1.** Physico-chemical parameters of raw effluent.

| S. No. | Samples          | Chromium      | Cadmium       | Zinc          | Iron          | Magnesium    |
|--------|------------------|---------------|---------------|---------------|---------------|--------------|
| 1      | Control water    | 0.24 ± 0.058  | 0.23 ± 0.044  | 2.96 ± 0.159  | 7.15 ± 0.269  | 1.87 ± 0.221 |
| 2      | Control plant    | ND            | 0.50 ± 0.076  | ND            | 1.18 ± 0.085  | 1.47 ± 0.139 |
| 3      | Raw effluent     | 10.65 ± 0.336 | 1.97 ± 0.21   | 1.69 ± 0.284  | 10.16 ± 0.187 | 0.70 ± 0.036 |
| 4      | Treated effluent | 0.62 ± 0.072  | 0.55 ± 0.086  | 0.855 ± 0.092 | 4.61 ± 0.367  | 0.29 ± 0.155 |
| 5      | Treated seeds    | 9.18 ± 0.099  | 1.40 ± 0.0717 | 0.685 ± 0.235 | 6.135 ± 0.648 | 1.22 ± 0.446 |
| 6      | F value          | 3662          | 340.97        | 37.39         | 129.6         | 13.96        |
| 7      | P value          | 0             | 0             | 0.0001        | 0.0001        | 0.0007       |

**Table 2.** Heavy metal analysis of treated effluent and treated *Strychnos potatorum* seeds.

| S. No. | Samples          | Chromium      | Cadmium      | Zinc         | Iron          | Magnesium    |
|--------|------------------|---------------|--------------|--------------|---------------|--------------|
| 1      | Control water    | 0.21 ± 0.176  | 0.21 ± 0.148 | 0.13 ± 0.053 | 6.03 ± 0.227  | 1.0 ± 0.078  |
| 2      | Control plant    | 1.53 ± 0.112  | 0.44 ± 0.127 | 2.03 ± 0.028 | 8.67 ± 0.415  | 1.31 ± 0.113 |
| 3      | Raw effluent     | 10.65 ± 0.336 | 1.97 ± 0.21  | 1.69 ± 0.284 | 10.16 ± 0.187 | 0.70 ± 0.036 |
| 4      | Treated effluent | 1.68 ± 0.091  | 1.87 ± 0.346 | ND           | 4.82 ± 0.264  | 0.4 ± 0.108  |

|   |               |              |              |              |             |              |
|---|---------------|--------------|--------------|--------------|-------------|--------------|
| 5 | Treated seeds | 8.34 ± 0.376 | 1.11 ± 0.096 | 2.32 ± 0.218 | 7.9 ± 0.224 | 1.24 ± 0.159 |
| 6 | F value       | 1160.26      | 22.7         | 405.24       | 746.18      | 119.99       |
| 7 | P value       | 0            | 0.0001       | 0            | 0           | 0            |

**Table 3.** Heavy metal analysis of treated effluent and treated *Moringa oleifera* seeds.

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| S. No. | Samples                       | Chromium (%) | Cadmium (%) | Zinc (%) | Iron (%) | Magnesium (%) |
|--------|-------------------------------|--------------|-------------|----------|----------|---------------|
| 1      | Effluent+ <i>S. Potatorum</i> | 94.17        | 72.08       | 49.7     | 54.62    | 58.57         |
| 2      | Effluent + <i>M. oleifera</i> | 84.22        | 5.07        | 100      | 52.55    | 42.85         |

**Table 4.** Biosorption % of Heavy metals by seeds.

| S. No. | Sample             | No. of Seeds | No. of Seeds Germinated | Germination % | Root length (cm) | Shoot Length(cm) | Seedling Vigour Index (VI) |
|--------|--------------------|--------------|-------------------------|---------------|------------------|------------------|----------------------------|
| 1.     | Control            | 30           | 30                      | 100           | 8.8 ± 0.765      | 2.9 ± 0.519      | 1210                       |
| 2.     | Water with seeds   | 30           | 27                      | 90            | 7.3 ± 0.643      | 2.4 ± 0.416      | 882                        |
| 3.     | Raw Effluent       | 30           | 12                      | 40            | 1 ± 0.288        | -                | 40                         |
| 4.     | Treated with seeds | 30           | 18                      | 60            | 3.3 ± 0.534      | 1 ± 0.165        | 258                        |

**Table 5.** Effect of *S. potatorum* seed treated effluent on the germination of *Vigna radiata* and seedling vigour index.

| S. No. | Sample             | No. of Seeds | No. of Seeds Germinated | Germination % | Root length (cm) | Shoot Length(cm) | Seedling Vigour Index (VI) |
|--------|--------------------|--------------|-------------------------|---------------|------------------|------------------|----------------------------|
| 1.     | Control            | 30           | 30                      | 100           | 8.8 ± 0.765      | 2.9 ± 0.519      | 1210                       |
| 2.     | Water with seeds   | 30           | 24                      | 80            | 7.1 ± 0.504      | 2.3 ± 0.63       | 882                        |
| 3.     | Raw Effluent       | 30           | 12                      | 40            | 1 ± 0.288        | -                | 40                         |
| 4.     | Treated with seeds | 30           | 18                      | 60            | 3.3 ± 0.606      | 0.5 ± 0.2        | 228                        |

**Table 6.** Effect of *M. oleifera* seed treated effluent on the germination of *Vigna radiata* and Seedling vigour index.

## Bioassay

Bioassay is an analytical method to determine the residual toxicity level of the treated effluent using living cells or tissues. The toxicity of raw effluent and treated effluent was estimated by germinating *Vigna radiata* seeds. The shoot and root length were measured, germination percentage and seedling vigour index was calculated (Tables 5 and 6).

In this present investigation the powdered seeds of *S. potatorum* and *M. oleifera* were used to biosorb the heavy metals from the tannery effluent. This method was effective since the concentration of Cr, Cd, Zn, Fe and Mg reduced from the tannery effluent (Tables 2 and 3) and was biosorbed by the seed powder (Table 4). The *S. potatorum* seeds contain Strychnine a crystalline alkaloid which is responsible for coagulation properties (Radin et al., 2014)

*M. oleifera* contain cationic protein that can clarify turbid water. They are reactive in high pH (4-6) due to salting mechanism of protein. Seedling vigour index increased in treated effluent when compared to raw effluent. The raw effluent contains higher concentration of the heavy metals which is deterrent to the germination and after the treatment metal ion concentration reduced in the effluent which enhances the germination percentage and seedling vigour index of the seeds. The tannery effluent after treating with *S. potatorum* and *M. oleifera* could be

used for watering the plants. Many researchers have reported on *S. potatorum* and *M. oleifera* seeds as ecofriendly and cheaper water purifier (Olsen 1987; Muthukumaran et al., 2013; Tripathi et al. 1976). Meneghel et al., 2013 stated that *M. oleifera* is biodegradable, non-toxic, non-corrosive and easy to use. *S. potatorum* was found to be environmentally friendly coagulants, which presented as a viable alternative for the treatment of wastewater and turbid water (Deshmukh et al., 2013). *M. oleifera* has potential as a natural Chromium removal agent that can be adopted by developing countries at low cost (Camilla, 2014).

## CONCLUSION

This research finding on biosorption ability of seeds like *S. potatorum* and *M. oleifera* in the treatment of tannery effluent was more effective. The tannery effluent after treating with these seeds could be used for irrigation purpose after investigating if there is any bioaccumulation of heavy metals in crop plants. The seed powder after treatment could be burnt and the adsorbed metals could be re-extracted. These seeds are a promising candidate for industrial effluent treatment to make water treatment process easier and environment friendly and it could replace the chemical coagulant.

## REFERENCES

- Abdulsalam, S., Mohammed, J., Olubiyo, S.E., and Tenimu, I.I. (2014). Biosorption of chromium (VI) Ion from tannery wastewater using two novel agricultural by-Products. *Int. J. Innov. Manag. AndTech.* 5(2): 78-82.
- Anderssonoch, C., Robertsson, J. (2014). Strychnos potatorum seed powder and fly ash. *Inter. J. Sci. Eng and Tech.Res.* 4(1):118-127.
- 2270
- Ndabigengesere, A. (1995). A study of the mechanism of coagulation optimization - flocculation of the *Moringa Oleifera*. These de
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- Characterization and removal of heavy metals in tannery effluents through the use of local materials. Tropical Ecology Working Group, Uppsala University, Sweden, ISSN 1654-5634.
- Deshmukh, B.S., Pimpalkar, S.N., Rakhunde, R.M., and Joshi, V.A. (2013). Evaluation performance of natural *Strychnos potatorum* over the synthetic coagulant alum, for the treatment of turbid water. *Int J Innov Res Sci Eng Technol.* 2(11): 6183-6189.
- Folkard, G. and Sutherland, J. (2002). Development of a naturally derived coagulant for water and wastewater treatment. *Wat Suppl.* 2(5-6): 89-94.
- Fourest, E. and Roux, C. J. (1992). Heavy metal biosorption by fungal mycelial by-products mechanisms and influence of pH. *Appl Microbiol Biot.* 38(3):399-403.
- Gatew, S., Mersha, W. (2013). Tannery waste water treatment using *Moringa stenopetala* seed powder extract. *Wyno Academic J. Phys. Sci.* 1(1): 1-8.
- Hong, A.H., Burmamu, B.R., Umaru, A.B., and Sadiq, U.M. (2017). Removal of heavy metals from contaminated water using *Moringa oleifera* seed coagulant in Yola and its environs. *Int. J. Eng. Invention.* 6(11): 40-45.
- Jayaram, K., Murthy, I.Y.N., Lalhruaitluanga, H., Prasad, M.N.V. (2009). Biosorption of lead from aqueous solution by seed powder of *Strychnos potatorum* L. *Colloids Surf B Biointerfaces.* 71(2): 248-254.
- Joshua, R. and Vasu, V. (2013). Characteristics of stored rain water and its treatment technology using *Moringa* seed. *Inter. J Life Sc. Bt & Pharm. Res.* 2(1): 155-174.
- Meneghel, A.P., Goncalve, A.C., Jr. Fernanda, R., Cardoso D.D., Antonio, C., et. al. (2013). Biosorption of cadmium from waste water using *Moringa (Moringa oleifera Lam.)* seeds. *Water Air Soil Pollut* 224:1383.
- Muthukumaran, G., Sasikala, S. Prakash, N. (2013). Proteins from natural coagulant for potential application of turbidity removal in water. *Inter J Eng and innovative Tech.* 3(1): 278-283.
- Muthuraman, G., and Sasikala, S. (2015). Kinetic studies for Chromium (VI) removal by using
- Doctorat, Universite de Sherbrooke, Sherbrooke, 202 p.
- Nduka Joseph, O., Michael, H., and Obuzor, G.U. (2015). Preliminary investigation on the extraction of heavy metals from produced water using *Moringa oleifera* leaves and seeds as adsorbents. *Res. J. Chem. Sci.* 5(1): 7-11.
- Olsen, A. (1987). Low technology water purification by bentonite clay and *Moringa oleifera* seed flocculation as performed in Sudanese village: effects on *Schistosomamansonii cercariae*. *Water Resources.* 21(5):517-522.
- Radin Mohamed, R.M.S., Abdul Rahman, N. and Mohdkassim, A.H. (2014). *Moringa oleifera* and *Strychnos potatorum* seeds as Natural coagulant compared with synthetic common coagulants in treating car wash wastewater : Case study1. *Asian J. Appl. Sci.* 2(5):693-700.
- Reddy, D.H.K., Lee, S.M., Sessaiah, K. (2012). Removal of Cd(II) and Cu(II) from aqueous solution by agro biomass: Equilibrium, kinetic and thermodynamic studies. *Environ. Eng. Res.* 17(3): 125-132.
- Senthil Kumar, P., Senthamarai, C., Sai Deepthi, A.S.L., Bharani, R. (2013). Adsorption isotherms, kinetics and mechanism of Pb(II) ions removal from aqueous solution using chemically modified agricultural waste. *Can. J. Chem. Eng.* 9999: 1-7.
- Senthilkumar, P., Rangasamy, G., Senthamarai, C., Priyadharshini, M., Anne Fernando, P.S., Ramakrishnan, S., Vaidhyanathan, V.K. (2012). Kinetics, mechanism, isotherm, and thermodynamic analysis of adsorption of cadmium ions by surface-modified *Strychnos potatorum* seeds. *Korean J. Chem. Eng.* 29(12): 1752-1760.
- Sutherland, J.P., Folkard, G.K. and Grant, W.D. (1990). Natural coagulants for appropriate water Treatment: A novel approach. *I Waterlines.* 8(4): 30-32.
- Tripathi, P.N., Chaudhuri, M. and Bokil, S.D. (1976). Nirmali seed: A naturally occurring coagulant. *Indian J. Environ. Health.* 18(4): 272-281.